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## **RESEARCH ARTICLE**

# Genetics of Yield and Fruit Quality Traits in Bitter Gourd (Momordica charantia L.)

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## **Abstract**

Utilization of genetic resources in crop breeding requires a basic understanding of the economic traits they are recognized for. Determination of genetics of traits helps to decide the appropriate breeding strategy for developing improved varieties. Segregating populations developed using *Momordica charantia* var. *charantia* and *M. charantia* var. *muricate* genotypes were used for generation mean analysis (GMA) to work out the gene effects controlling yield and fruit quality traits in bitter gourd. The study confirmed the role of non-allelic interactions and predominance of dominance gene effect (h) for most economic traits, which indicates the suitability of recurrent selection or heterosis breeding for the improvement of these traits. Segregation analysis indicated the single dominant gene control of fruit epicarp color, tubercle nature and fruit ridges. The predominance of complementary type of epistasis for most traits implies the genetic divergence between the parents used and the possibility of realizing a greater genetic gain in the breeding programme using these genetic resources.

Keywords: Dominance effect, Epistasis interaction, Generation mean analysis, Genetic studies, Momordica charantia.

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# Introduction

Momordica charantia L. (Syn. bitter gourd, balsam pear) (2n = 22) is a traditional vegetable and medicinal crop of tropics and subtropics of Asia, South America and East Africa (Heiser, 2016). Bitter gourd belongs to Cucurbitaceae family and is known for its distinctive bitter flavor and high nutritional value, specifically ascorbic acid and iron (Behera, 2004). As a medicinal crop, it is widely known for its anti-diabetic, anti-HIV and anti-tumor properties (Behera et al., 2008). Bitter gourd is a highly cross-pollinated crop due to its monoecious sex form, which results in wide variability. Mainly, Indian bitter gourd gene pool exhibits high morphological variation in growth habit, maturity, fruit shape, size, color, surface texture and sex expression (Behera et al., 2006). This genetic variability is vital in developing varieties as per consumer demand. Market preference for bitter gourd varieties varies from region to region with respect to its fruit quality traits, such as fruit color, length, shape, and tubercles (Rathod et al., 2019). For instance, dark green to glossy green fruits are preferred in northern India. White fruits are ideal in the south Indian market, whereas in eastern parts of the country, small and dark green fruits are desired (Behera et al., 2010). Hence to exploit the existing genetic variation in order to develop varieties suitable for various market segments, elucidating genetics of these traits is crucial. To successfully determine trait genetics, populations derived from diverse parents should be used for genetic analysis. The generated knowledge through this

Table 1: Generation means for earliness and yield traits in cross PVGy-201 × PDM and CBM-12 × DBG-4 of bitter gourd

Traits	Cross	P1	P2	F1	F2	B1	B2
Node to first	PVGy-201 × PDM	3.05 ± 0.29	11.400 ± 0.72	5.62 ± 0.36	6.57 ± 0.16	$4.83 \pm 0.23$	9.78 ± 0.31
female flower	CBM-12 $\times$ DBG-4	$12.60 \pm 0.77$	$7.400 \pm 0.64$	$7.82 \pm 0.39$	$7.95 \pm 0.40$	$9.91 \pm 0.26$	$6.01 \pm 0.28$
Days to first	$PVGy-201 \times PDM$	$26.80 \pm 0.57$	45.250 ± 1.01	$32.10 \pm 0.38$	$35.17 \pm 0.46$	$30.36 \pm 0.32$	$36.90 \pm 4.33$
female flower	CBM-12 $\times$ DBG-4	$79.45 \pm 1.08$	$78.450 \pm 1.60$	$63.450 \pm 1.60$	$71.34 \pm 0.68$	$69.46 \pm 0.48$	$66.33 \pm 0.74$
Node to first	PVGy-201 $\times$ PDM	-	-	-	-	-	-
male flower	CBM-12 $\times$ DBG-4	$9.05 \pm 0.39$	$4.15 \pm 0.38$	$4.45 \pm 0.26$	$7.12 \pm 0.33$	$6.30 \pm 0.20$	$4.16 \pm 0.17$
Days to first	$PVGy-201 \times PDM$	-	-	-	-	-	-
male flower	CBM-12 $\times$ DBG-4	$64.20 \pm 1.34$	65.95 ± 1.39	$55.82 \pm 0.89$	$68.67 \pm 0.80$	$61.66 \pm 0.62$	$66.33 \pm 0.74$
Fruit length	$PVGy-201 \times PDM$	$113.87 \pm 3.30$	$180.39 \pm 2.27$	$165.48 \pm 1.78$	$144.30 \pm 2.71$	140.53 ± 1.42	$170.78 \pm 1.39$
(mm)	CBM-12 $\times$ DBG-4	54.19 ± 1.16	165.35 ± 1.90	$109 \pm 0.90$	99.52 ± 1.79	$73.01 \pm 0.83$	$128.62 \pm 0.91$
Fruit weight	PVGy-201 $\times$ PDM	89.47 ± 4.26	145.27 ± 2.89	139.84 ± 1.352	120.46 ± 2.30	128.29 ± 1.12	147.39 ± 1.15
(gm)	CBM-12 $\times$ DBG-4	$17.38 \pm 0.68$	96.85 ± 1.79	$56.65 \pm 0.93$	47.32 ± 1.65	$29.89 \pm 0.60$	$70.25 \pm 0.92$
Fruit diameter	$PVGy-201 \times PDM$	$42.82 \pm 0.81$	$38.97 \pm 0.72$	$44.46 \pm 0.57$	$40.79 \pm 0.35$	$42.91 \pm 0.45$	$38.06 \pm 0.46$
(mm)	CBM-12 $\times$ DBG-4	$34.56 \pm 0.94$	$41.55 \pm 0.91$	$38.12 \pm 0.44$	$38.49 \pm 0.53$	$34.83 \pm 0.35$	$39.483 \pm 0.29$
Number of	$PVGy-201 \times PDM$	$20.70 \pm 0.84$	$13.20 \pm 0.56$	$22.52 \pm 0.64$	19.01 ± 0.45	$23.95 \pm 0.51$	$18.05 \pm 0.57$
fruits per plant	CBM-12 $\times$ DBG-4	$24.30 \pm 1.54$	$13.50 \pm 1.03$	$25.37 \pm 0.87$	$18.47 \pm 0.50$	$19.85 \pm 0.61$	$16.33 \pm 0.43$
Fruit yield per	PVGy-201 × PDM	1,505.0 ± 100.9	1,775.7 ± 97.9	2,746.5 ± 94.1	1,556.2 ± 61.2	2,682.7 ± 74.9	2,261.1 ± 84.1
plant (gm)	CBM-12 $\times$ DBG-4	431.7 ± 35.4	1,308.5 ± 105.7	1,427.7 ± 43.8	843.5 ± 35.8	682.9 ± 22.5	1,288.1 ± 38.3

Table 2: Test of significance of A, B, C, and D scales earliness and yield traits in cross PVGy-201 × PDM and CBM-12 × DBG-4 of bitter gourd

Traits	Cross	Α	В	С	D
Node to first female	PVGy-201 × PDM	-1.09* ± 0.46	-2.54* ± 0.72	3.39* ± 0.88	-3.51** ± 0.36
flower	CBM-12 × DBG-4	$0.59 \pm 0.72$	$3.19* \pm 0.66$	4.2* ± 1.447	$-0.22 \pm 0.62$
Days to first female	PVGy-201 × PDM	-1.83* ± 0.67	3.55* ± 0.98	-4.468* ± 1.635	3.09** ± 0.75
flower	CBM-12 × DBG-4	3.96* ± 1.53	9.23** ± 1.91	-0.57 ± 3.278	6.88** ± 1.15
Node to first male	PVGy-201 × PDM	-	-	-	-
flower	CBM-12 × DBG-4	$0.90^* \pm 0.44$	$0.26 \pm 0.41$	-6.39** ± 1.08	3.77** ± 0.50
Days to first male	PVGy-201 × PDM	-	-		-
flower	CBM-12 × DBG-4	-3.30* ± 1.44	-10.89** ± 1.57	-32.90** ± 2.93	9.35** ± 7.06
F. Marcalle (com)	PVGy-201 × PDM	-1.71 ± 3.33	$4.302 \pm 2.84$	48.02** ± 8.57	-22.71** ± 4.09
Fruit length (mm)	CBM-12 × DBG-4	6.20** ± 1.59	6.15* ± 1.99	17.56* ± 5.49	$-2.60 \pm 2.68$
F 1./ .	PVGy-201 × PDM	-27.26** ± 3.53	-9.67* ± 2.79	32.56** ± 7.71	-34.75** ± 3.45
Fruit weight (gm)	CBM-12 × DBG-4	4.25** ± 1.18	2.98 ± 1.94	18.23** ± 5.04	-5.49* ± 2.46
F. H. H I ()	PVGy-201 × PDM	-1.53 ± 0.95	4.31** ± 0.92	1.54 ± 1.50	$0.62 \pm .067$
Fruit diameter (mm)	CBM-12 × DBG-4	3.01* ± 0.89	$0.71 \pm 0.832$	-1.61 ± 1.87	2.67* ± 0.82
Number of fruits	PVGy-201 × PDM	-4.67** ± 1.04	-0.37 ± 1.01	$2.89 \pm 1.73$	-3.97** ± 0.84
per plant	CBM-12 ×DBG-4	7.97** ± 1.52	4.20** ± 1.13	10.64** ± 2.29	$0.768 \pm 0.88$
Fruit yield per plant	PVGy-201 × PDM	-1,113.8** ± 144.0	-0.14 ± 152.9	2,548.6** ± 239.9	-1,831.3** ± 117.6
(gm)	CBM-12 × DBG-4	293.66** ± 5.75	-40.06 ± 0.41	821.63** ± 5.76	-284.01** ± 4.7

<sup>(\*, \*\*</sup> indicated significant effect at p = 5 and 1%)

study will help in better utilization of these germplasm in crop improvement through appropriate breeding methods.

Momordica charantia is the most widely grown species among the seven Momordica species found in India (Joseph & Antony 2010). Bitter gourd cultivars in India fall into two

botanical groups, *M. charantia var. charantia*, which produces large fusiform fruits and *M. charantia var. muricata* (wild), which bears small and disc-shaped fruit with numerous tubercles on its surface (Chakravarty 1990). These two recognized botanical varieties show striking morphological

**Table 3:** Estimates of genetic effects for earliness and yield traits in cross PVGv-201 × PDM and CBM-12 × DBG-4 of bitter gourd

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Traits	Cross	[m]	[d]	[h]	[1]	[j]		X2	Epistasis
Node to first	$PVGy-201 \times PDM$	$5.57^{**} \pm 0.11$	$4.90^{**} \pm 0.278$	$5.42^{**} \pm 0.818$	$7.02^{**} \pm 0.725$	$-1.45 \pm 0.784$	$-10.65^{**} \pm 1.423$	94.37**	D
female flower	CBM-12 $\times$ DBG-4	$7.85^{**} \pm 0.283$	$3.90^{**} \pm 0.274$	$-1.72 \pm 1.335$	$0.45 \pm 1.256$	$2.60* \pm 0.897$	$3.32^{**} \pm 1.815$	24.72*	U
Days to first	$PVGy-201 \times PDM$	$35.17^{**} \pm 0.325$	$-6.53^{**} \pm 0.382$	$-10.11^{**} \pm 1.588$	$-6.18^{**} \pm 1.509$	$5.38^{**} \pm 1.122$	$7.90* \pm 2.236$	38.69**	О
female flower	CBM-12 $\times$ DBG-4	$71.34^{**} \pm 0.484$	$3.13^{**} \pm 0.628$	$-29.27^{**} \pm 2.659$	$-13.77^{**} \pm 2.306$	$5.26* \pm 1.857$	$26.97^{**} \pm 4.129$	47.61**	U
Node to first male	Node to first male $PVGy-201 \times PDM$	ı	1	ı	1		1	,	ı
flower	CBM-12 $\times$ DBG-4	$7.12^{**} \pm 0.234$	$2.13^{**} \pm 0.188$	$-9.70*8 \pm 1.043$	-7.55** ± 1.008	$-0.63 \pm 0.541$	$8.72^{**} \pm 1.317$	57.01**	U
Days to first male	PVGy-201 ×PDM	ı	1	ı	1		1	,	ı
flower	CBM-12 $\times$ DBG-4	$68.67^{**} \pm 0.565$	-4.66** ± 0.688	$-27.95*8 \pm 2.807$	$-18.70^{**} \pm 2.647$	$-7.58* \pm 1.943$	$4.50 \pm 4.022$	140.0**	ı
(20,20) 4400 00 (11,11,11)	PVGy-201 ×PDM	$144.30^{**} \pm 1.92$	$-30.25^{**} \pm 1.40$	$63.77^{**} \pm 8.40$	$45.43^{**} \pm 8.18$	$6.01 \pm 4.0$	-42.84** ± 10.25	36.96**	U
rruit iengtii (mim)	CBM-12 $\times$ DBG-4	$99.52^{**} \pm 1.268$	$-55.60^{**} \pm 0.873$	$-6.52^{*} \pm 5.465$	$5.20 \pm 5.362$	$-0.04 \pm 2.355$	$7.16^{**} \pm 6.506$	25.38**	О
(	$PVGy-201 \times PDM$	$120.46^{**} \pm 1.63$	$-19.10^{**} \pm 1.140$	$91.98^{**} \pm 7.213$	$69.51^{**} \pm 6.913$	$17.59^{**} \pm 4.299$	-106.45** ± 8.96	148.4**	U
rruit weignt (gm)	CBM-12 $\times$ DBG-4	$47.32^{**} \pm 1.169$	$-40.36** \pm 0.784$	$0.52 \pm 5.024$	$10.99^* \pm 4.934$	$-1.26 \pm 2.076$	$-3.75 \pm 5.942$	21.92**	ı
Fruit diameter	$PVGy-201 \times PDM$	$40.79^{**} \pm 0.249$	4.85** ± 0.457	$-0.67 \pm 1.463$	$-1.24 \pm 1.351$	$5.85^{**} \pm 1.195$	$4.02 \pm 2.363$	28.30**	ı
(mm)	CBM-12 $\times$ DBG-4	$38.49^{**} \pm 0.377$	$-4.64^{*} \pm 0.326$	$-5.28^{**} \pm 1.737$	$-5.34^{**} \pm 1.644$	$-2.30^{*} \pm 1.136$	$9.072^{**} \pm 2.288$	18.84**	О
Number of fruits	$PVGy-201 \times PDM$	$19.01^{**} \pm 0.324$	$5.90^{**} \pm 0.549$	$13.52^{**} \pm 1.794$	$7.94^{**} \pm 1.699$	$4.30^{*} \pm 1.311$	$12.99^{**} \pm 2.799$	35.57**	U
per plant	CBM-12 $\times$ DBG-4	$18.47^{**} \pm 0.355$	$3.51^{**} \pm 0.534$	$2.94 \pm 1.992$	$-1.53 \pm 1.776$	$-3.76* \pm 1.695$	$13.71^{**} \pm 3.135$	35.11**	U
Fruit yield pe	$PVGy-201 \times PDM$	$1,556.2^{**} \pm 43$	421.5** ± 79.6	$4,768.7^{**} \pm 249$	$3,662.6^{**} \pm 235$	$1,113.7^{**} \pm 187$	$4,776.6** \pm 398$	297.5**	O
plant (gm)	CBM-12 × DBG-4	$843.53^{**} \pm 25.3$	$-605.2^{**} \pm 31.4$	$925.6^{**} \pm 129.4$	$568.0^{**} \pm 119.3$	$-333.7^{*} \pm 100.8$	$314.4^{**} \pm 190.1$	66.82**	U
(*, ** indicated sic	(*, ** indicated significant effect at $p = 5$ and 1%)	= 5 and 1%)							

differences and exhibit remarkable diversity for economic traits. Hence, using these two botanical types in breeding programs is essential for effective bitter gourd improvement.

Yield is a complex polygenic trait, determined by the cumulative effects of several yield-contributing traits such as fruit length, weight, number of female flowers, etc. Earliness is another important trait that is directly correlated with yield. Farmers prefer early varieties due to its suitability to multiple cropping systems and ability to fetch high prices in the early season market (Dey et al., 2012). To breed highyielding early varieties, basic understanding of the genetics of component traits and knowledge of gene action/effects functioning in a particular breeding population is essential (Vinay et al., 2023). Keeping above facts in mind, current generation mean analysis study was conducted to estimate the various genetic components of economic traits in bitter gourd using population derived from genetically diverse parental lines. The results of this study will lay the foundation for development of novel early-high yielding varieties and hybrids of bitter gourd to boost the production.

#### **Material and Methods**

Present investigation was conducted at experimental field of ICAR-Indian Agricultural Research Institute, New Delhi, India. For genetic analysis, two cross combinations PVGy-1 × Pusa Do Mausami (PDM) and CBM-12 × DBG-4, were used to develop populations (F1, F2, B1 and B). PVGy1, PDM and DBG-4 lines belong to M. charantia var charantia where as CBM-12 belongs to M. charantia var muricata group. The F, progenies were developed during rainy season (June-October, 2018), F<sub>2</sub> and backcross progenies (B<sub>1</sub> and B<sub>2</sub>) were genertaed from F, plants during the spring-summer of 2019 under the insect-proof net house. Final trial for evaluation was laid out with six generations  $(P_1, P_2, F_1, F_2, B_1 \text{ and } B_2)$  under natural/open field conditions during August- October of 2019 in a completely randomized block design with three replications. The data was recorded from 10 plants in each parent, 20 plants of F<sub>1</sub>, 30 plants of B<sub>1</sub> and B<sub>2</sub> generations and 118 (PVGy-1 × Pusa Do Mausami) and 102 (CBM-12 × DBG-4) plants in F, progenies. The phenotypic observations were recorded for four earliness-related traits, namely, node to first female flower (NFFF), days to first female flower (DFFF), node to first male flower (NFMF) and days to first male flower (DFMF) and five yield traits viz. namely, fruit length (cm), fruit diameter (cm), fruit weight (g), number of fruits per plant and fruit yield/plant (g). Two male flower-related parameters (NFMF, DFMF) were not considered in first cross (PVGy-1 × PDM) since the female parent (PVGy-1) was a gynioecious line. Spacing, nutrition, irrigation, cultural practices and plant protection measures were followed as per recommendations to raise a successful crop in the experimental plots.

Generation mean analysis (GMA) was performed as suggested by Hayman (1958) to estimate genetic

 $\textbf{Table 4:} Inheritance\ pattern\ of\ fruit\ epicarp\ color,\ tubercle\ nature\ and\ ridgeness\ in\ bitter\ gourd\ in\ PVGy-201\times PDM\ and\ CBM-12\times DBG-4\ of\ bitter\ gourd$ 

bitter gourd						
		Fruit epi	carp colour in cros	s - CBM-12 × DBG-4		
Population	Total	Observed	d	Expected ratio	χ2 value	p-value
	Green fruits	White fro	uits			
P1(CDM-12)	10	10	-	-		
P2 (DBG-4)	10	-	10	-		
F1(CBM-12 $\times$ DBG-4)	20	20	-	-		
F2	102	80	22	3:1	0.64	0.43
B1(F1 $\times$ CBM-12)	30	30	-	-		
$B2(F1 \times DBG-4)$	30	17	13	1:1	0.53	0.47
		Fruit tub	ercle nature in cro	ss - CBM-12 × DBG-4		
Population	Total	Observed	d	Expected ratio	χ2 value	p-value
	Conspicuous	Non-con	spicuous			
P1(CDM-12)	10	10	-	-		
P2 (DBG-4)	10	-	10	-		
F1(CBM-12 $\times$ DBG-4)	20	20	-	-		
F2	102	74	28	3:1	0.33	0.57
B1(F1 $\times$ CBM-12)	30	30	-	-		
$B2(F1 \times DBG-4)$	30	16	14	1:1	0.13	0.72
	Fruit	ridgeness i	n cross - PVGy-201	× Pusa Do Mausami (PDM)		
Population	Total	Observed	d	Expected ratio	χ2 value	p-value
	Dis-continuous	Continue	ous			
P1(PVGy-201)	10	10	-	-		
P2 (PDM)	10	-	10	-		
F1(PVGy-1 $\times$ PDM)	20	20	-	-		
F2	118	84	34	3:1	0.92	0.34
B1(F1 × PVGy- 201)	30	17	13	1:1	0.53	0.47
$B2(F1 \times PDM)$	30	30	-	-		

components. At first, using means of different generations, ABCD scales (Hayman and Mather, 1955) were tested to check for the presence of non-allelic interactions (epistasis) and then the data was subjected to joint scaling test which integrates multiple scaling tests to examine the competence of simple additive-dominance model or to detect epistasis using  $\chi 2$  test. The six-parameter or di-genic interaction model (Hayman, 1958) was used to estimate the gene effects whenever the  $\chi 2$  and ABCD scaling test is inadequate. These parameters represent mean effect [m], genetic effects including additive [d] and dominance [h], and gene interaction effects comprising additive × additive [i], additive × dominance [j] and dominance × dominance [l]. Student 't' test was used to declare the significance of the estimated scales (ABCD) and gene effects (m, d, h, i, j, l). The type of epistasis was determined as per Kearsey and Pooni, 1996.

To study the genetic inheritance of fruit traits viz. fruit epicarp color, ridgeness, tubercle nature, chi-square ( $\chi$ 2) test suggested by Panse and Sukhatme (1985) was done. These traits are expected as controlled by single gene.

## **Results and Discussion**

The mean values (Table 1) of F, for fruit length, fruit weight and fruit diameter (in CBM-12 × DBG-4) were greater than the mid-parent values (average of the two parents) which indicate average heterosis, whereas F, mean values of number of fruits per plant and yield per plant and fruit diameter (in PVGy-201 × PDM) was higher than better parent values which indicated over dominance/ heterobeltotis. Similarly, for earliness-related traits, in both cross combinations mean values of F, implied negative average heterosis which is desirable. These results suggest the scope for improvement yield and earliness through heterosis breeding. There was decline in mean values of yield traits of F<sub>2</sub> population than corresponding F<sub>1</sub> in all yield traits. This apparently indicated influence of inbreeding depression. These findings are in agreement with the results obtained by Dey et al. (2012) in bitter gourd. The mean effect (m) was significant for all studied traits among three crosses, indicated that the traits were quantitatively inherited.

Significance of one or more scales (A, B, C, D) as shown in Table 2 and joint scaling  $\chi 2$  (Table 3) test for all the traits under study in both crosses implies presence of digenic non-allelic interaction (epistasis). Hence, the simple additive-dominance model cannot satisfactorily explain the genetics of these traits. Therefore, according to Jinks and Jones (1958), the six-parameter model was employed to estimate six components of genetic variation, viz. m, d, h, i, j and l. The presence of non-allelic gene interactions involved in genetic control of quantitative yield traits in bitter gourd was previously reported by Kumari et al. (2015).

Higher magnitude and highly significant values of dominance effect (h) in desired direction was obtained for earliness-related traits like days to first female flower (both cross), node to first male flower (CBM12  $\times$  DBG-4) and day to first male flower (CBM12  $\times$  DBG-4). Additive  $\times$  additive (i) type of non-allelic interaction was highly significant and negative for these traits. Similarly, higher magnitude significant-positive values of 'h' was observed for major yield parameters, number of fruits per plant and fruit yield per plant in both crosses, which indicates predominance of dominant gene effect. To add to this, of the three non-allelic interactions dominance × dominance (I) gene interaction was found to be positive and significant for these yield related traits. Hence, while breeding for improvement of earliness and yield traits selection should be delayed until heterozygosity is reduced in population. According to result of our study, hybrid development through heterosis breeding would be the most appropriate breeding method to evolve early and high-yielding hybrids using these cross combinations. For fruit length and fruit weight traits in cross PVGy-201  $\times$  PDM, additive effect (d) was positively significant with higher magnitude than dominance effect (h). In addition to this, additive × additive (i) type of non-allelic interactions was found to be highly significant. This clearly implies the suitability of simple selection procedure for the improvement of these traits. Complementary epistasis was observed for most earliness and yield traits under study, which implies that parents selected for crossing were diverse for that particular trait and desirable alleles for the traits are distributed among the parents. Hence, it is possible to realize greater genetic gain in a breeding programme. Our results are supported by the findings of Dey et al. (2012) in Bitter gourd.

The inheritance pattern of fruit epicarp color and tubercle nature was studied using cross of CBM-12 (green fruits with conspicuous tubercle) × DBGS-4 (white fruit with non-conspicuous tubercle). The analyzed result presented in Table 4 implies monogenic nature of both traits. Green fruit epicarp color is dominant over white, and conspicuous tubercles nature was dominant over non-conspicuous. Similar monogenic dominant nature of inheritance of fruit color was earlier reported by Hu *et al.* (2002) in bitter gourd. Our result of genetics of tubercle nature is supported by

earlier studies (Rao *et al.*, 2022). Phenotypic data obtained from cross PVGy-201 (dis-continuous) × PDM (continuous) was utilized to study the genetics of fruit ridgeness. The segregation pattern of fruit ridgeness in filial and backcross generations supported the single dominant gene hypothesis of genetic control. Discontinuous ridgeness was dominant over continuous ridgeness. Our study's result is supported by Rao *et al.*'s findings, 2022.

## Conclusion

The genetic analysis for estimation of gene effects controlling economic traits in bitter gourd was done using diverse parental lines. Both additive and non-additive gene interactions control the economically important earliness and yield traits in bitter gourd. Therefore, recombination breeding with selection in later generations would be the most suitable way to improve these quantitative traits. Simple selection can give desired results when additive gene action is predominant. At the same time, dominant gene effects should be concentrated for better results. The traits governed by additive gene effect can be effectively improved through pedigree method. On the other hand, traits regulated by non-additive gene interaction (dominance effect) could be improved by heterosis breeding and through recurrent selection for specific combining ability. The traits with high magnitude of dominance and additives gene effects can be improved by pedigree or bulk methods or reciprocal recurrent selection methods. From the present study and most of the earlier reports, it is now clear that the fruit quality traits (color, tubercles and ridgeness) are qualitatively inherited i.e. by single dominant gene. Simple genetics of these fruit traits will permit the bitter gourd breeder to incorporate these traits in hybrids. The predominance of complementary epistasis for most economic traits studied implies the genetic divergence between the parents used and the possibility to realize greater genetic gain in breeding programme using these genetic resources. The study also implies the importance of populations derived from diverse genetic resources for the effective genetic analysis of phenotypic expressions in bitter gourd.

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